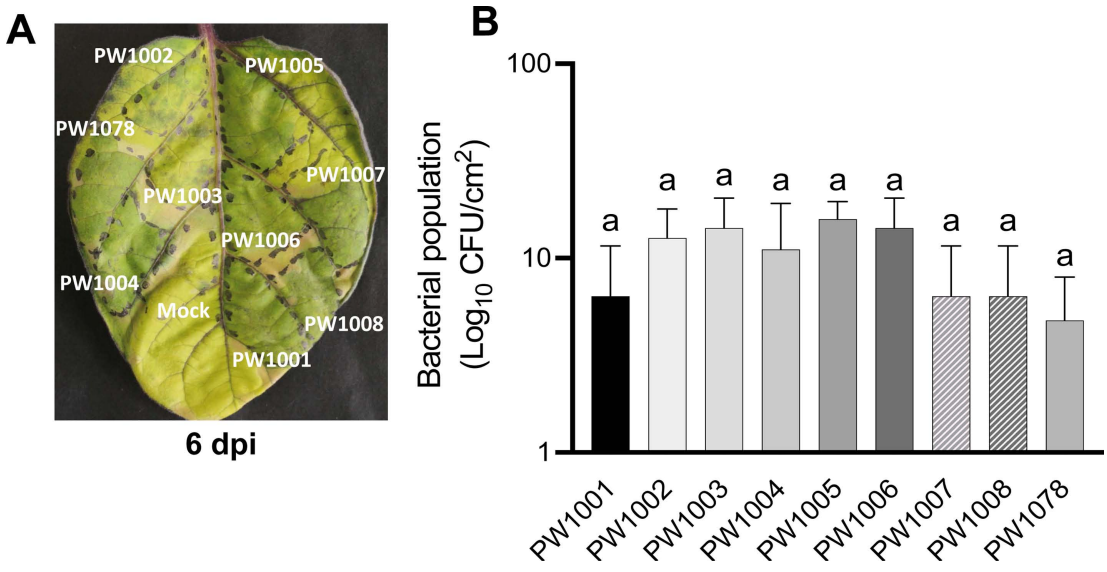
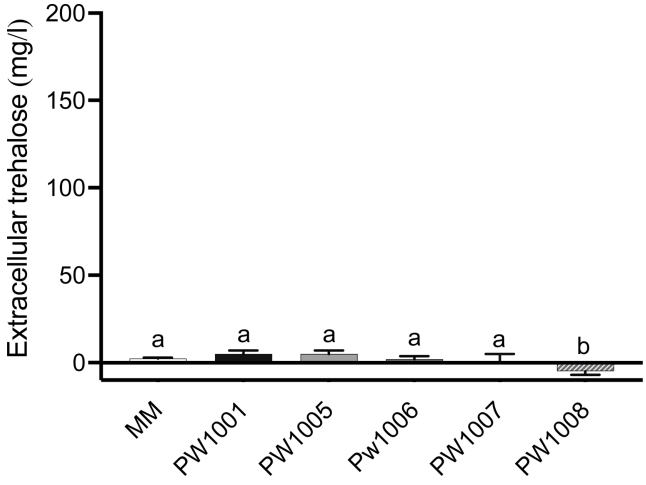
Supplementary Table 1. Primers used in this study

Primer name	Sequence (5′–3′)	Purpose
P3747	ctctagaggatccccGACCAGCTGCGCGTGGTGTCGTG	PCR to amplify 0.8 kb of *hrpB* region for cloning to generate pARO::*hrpB*
P3748	tcgagctcggtacccGGTCGCTGCGCTCGATGTTCTCG	
P3749	ctctagaggatccccCTGCACGGCGGCATGGACGGCGC	PCR to amplify 0.8 kb of *hrcV* region for cloning to generate pARO::*hrcV*
P3750	tcgagctcggtacccCACGACGTGATCGGCGATCACGC	
hpaB-F	GGTCTCACAAGAGGCCACTT	PCR to amplify 1,911 bp of *hpaB* and surrounding region for cloning
hpaB-R	GCGATGAGAACAGCAGCCAT	
hpaB-d1	ggctctagaCGCGTTGCTCAAGACTCATG	Inverse PCR to delete *hpaB* coding region to generate Δ*hpaB*
hpaB-d2	ggctctagaCGCATGATGGCGTCTTCTAA	
ripTPS-F	TTGCGGAGTGTTGAGGTTGC	PCR to amplify 2,958 bp of *ripTPS* and surrounding region for cloning
ripTPS-R	CTGCGTTCCAGATCATCAAC	
ripTPS-d1	ggctctagaCGTCGTGCTTCTGGTGGTAT	Inverse PCR to delete *ripTPS* coding region to generate Δ*ripTPS*
ripTPS-d2	ggctctagaCGCGTCTATGGATGTGTGAC	
otsA-F_IF	cggtacccggggatcGCTCAAGGCGCTGGTGCAGCCCG	PCR to amplify 678 bp of *otsA* region for cloning to generate pARO::*otsA*
otsA-R_IF	cgactctagaggatcTGCTTGTTGATGTAGCGGATCGG	
otsB-F_NEB	gtcacactgcttccggaattcATGGCACGCTCGTTGACATC	PCR to amplify 600 bp of *otsB* region for cloning to generate pARO::*otsB*
otsB-R_NEB	acaggtgcggttgctgaattcGTGCCCACCTTGATCGACCA	
otsB-F	CGTATCCTCGCGAGCGTATC	PCR to amplify *otsBA* region for complementation
otsA-R	GCGCTATCTAATAGCAAGCG	
otsA-1	GAGCCGATTGATCGTGGTCTC	Colony PCR in pair with M13RV to confirm pARO::*otsA* construction
otsB-1	TTGCCTCAGATCGCGTCACC	Colony PCR in pair with M13RV to confirm pARO::*otsB* construction
ripX-F	ggtaccCGTTTTCATGACCCGGCTTC	PCR to amplify 449 bp of *ripX* and promoter region for cloning to generate pHRP309::*ripX*
ripX-R	gcggccgcGATCAGGTCTTGCACGGACT	
ripX-F2	tctagaCGTTTTCATGACCCGGCTTC	PCR to amplify 1,324 bp of *ripX* and promoter region with ripX-F to generate pARO-*ripX*::'*cya*
ripX-R2	ctcgagCATCGGCTGCGTCGTGGTGT	

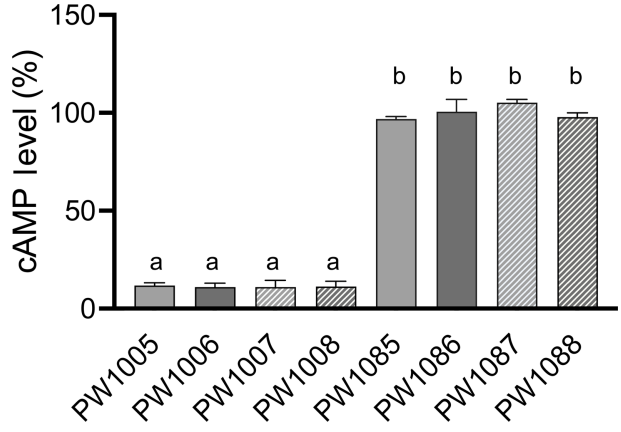
Small letters indicate additive nucleotides for cloning of *hrpB*, *hrcV*, *otsA* and *otsB* fragments. Underlined letters indicate artificial *Eco*RI sites in otsB-F_NEB and otsB-R_NEB, and *Xba*I sites in hpaB-d1, hpaB-d2, ripTPS-d1, ripTPS-d2 and ripX-F2, *Kpn*I site in ripX-F, *Xho*I site in ripX-R2, and *Not*I site in ripX-R.



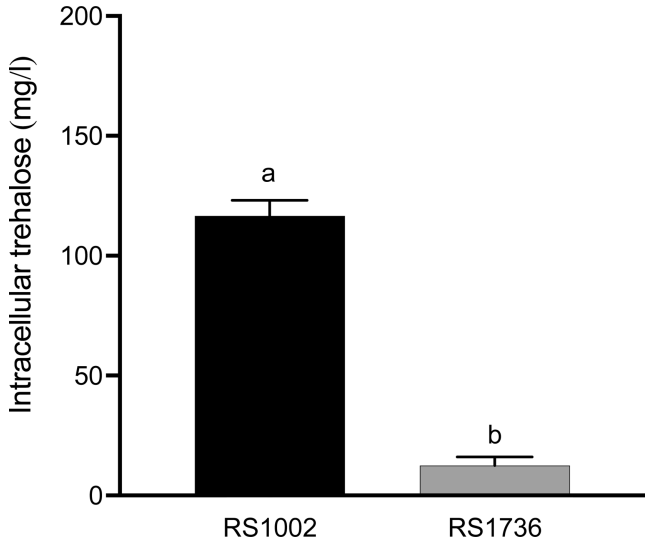
Supplementary Fig. 1. Assessment of virulence of *Ralstonia syzygii* subsp. *indonesiensis* (*Rsi*) PW1001 on eggplant *Solanum melongena* cv. Senryo-nigou. (A) Leaf infiltrated with the bacterial suspension (OD_{600} of 0.0003) of *Rsi* PW1001 (WT), PW1002 (*hrpB*), PW1003 (*hrcV*), PW1004 (Δ*hpaB*), PW1005 (*otsA*), PW1006 (*otsB*), the complemented strains PW1007 (*otsA* Tn*otsBA*$^+$) and PW1008 (*otsB* Tn*otsBA*$^+$), PW1078 (Δ*ripTPS*) and 10 mM $MgSO_4$ (Mock). Photograph shows a representative results from two independent experiments with similar results at 6 dpi. (B) The bacterial population was determined at 6 dpi. Values are means ± SD of two replicates from two independent experiments. The same letters indicate no significant differences among the nine inoculations as defined by Tukey's multiple comparison test ($P < 0.05$).



Supplementary Fig. 2. Trehalose concentration in the spent medium of each bacterial culture. Trehalose concentration was measured by Trehalose Assay Kit. Values are means ± SD of three replicates. Different letters indicate significant differences among the five strains and negative control MM (minimum medium) as defined by Tukey's multiple comparison test ($P < 0.05$).



Supplementary Fig. 3. Translocation of RipX effector. The cAMP levels of eggplant *Solanum melongena* cv. Senryo-nigou leaves inoculated with *Rsi* PW1005 (*otsA*), PW1006 (*otsB*), PW1007 (*otsA* Tn*otsBA*$^+$) and PW1008 (*otsB* Tn*otsBA*$^+$), PW1085 (*otsA ripX*::'*cya*), PW1086 (*otsB ripX*::'*cya*), PW1087 (*otsA TnotsBA*$^+$ *ripX*::'*cya*), and PW1088 (*otsB TnotsBA*$^+$ *ripX*::'*cya*). Values are means ± SD of two replicates from two independent experiments. Different letters indicate significant differences among the eight strains as defined by Tukey's multiple comparison test ($P < 0.05$).



Supplementary Fig. 4. Trehalose concentration of trehalose samples prepared from bacterial cells, RS1002 and RS1736, that were cultured in minimal medium for 16 h. Trehalose concentration was measured by Trehalose Assay Kit. Values are means ± SD of three replicates from two independent experiments. Different letters indicate significant differences among two strains as defined by Tukey's multiple comparison test ($P < 0.05$).