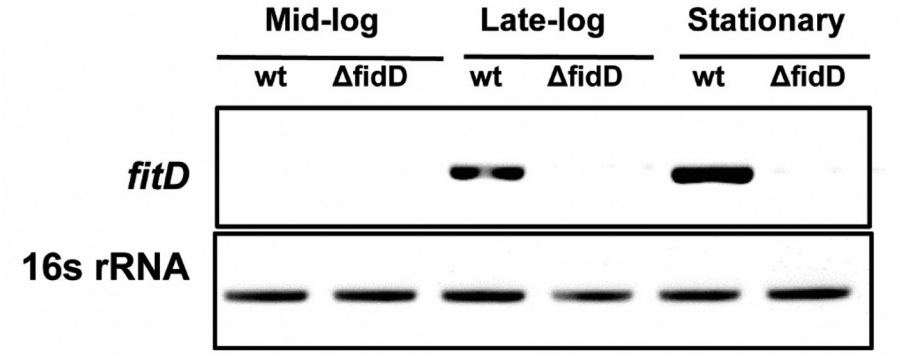
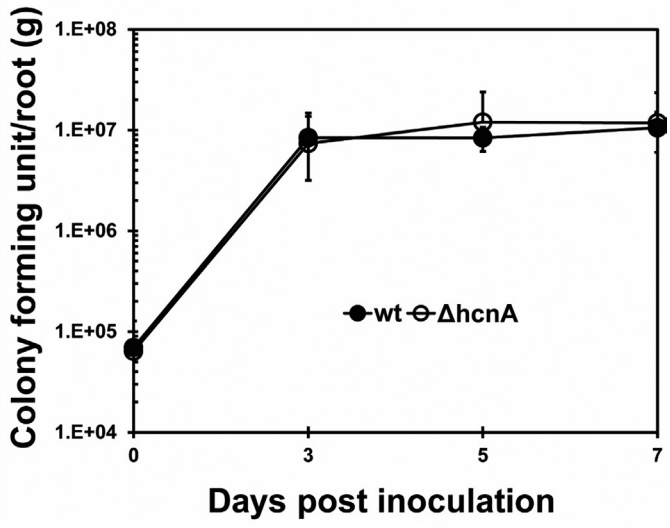
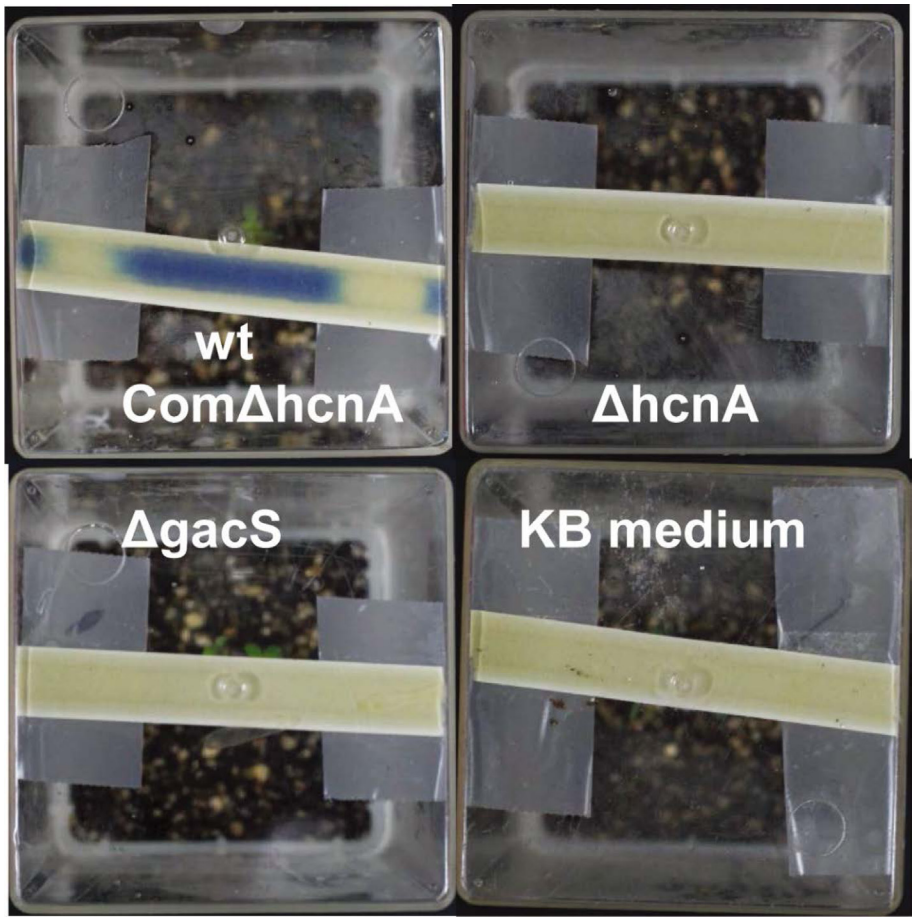


Supplementary Fig. 1. Transcript analysis of the *fitD* gene in *Pseudomonas chlororaphis* O6 wild-type (wt) and *fitD* mutants (Δ*fitD*). Cells grown in KB broth were harvested in the mid-logarithmic phase ($OD_{600\ nm}$ = 0.8), late-logarithmic phase ($OD_{600\ nm}$ = 1.8), and stationary phase ($OD_{600\ nm}$ = 2.4). Total RNA was isolated by the RNeasy Mini Kit (QIAGEN GmbH, Hilden, Germany) and 200 ng was added to the reactions established following the instructions provided with the QuantiTect SYBR Green reverse transcription PCR kit (QIAGEN GmbH, Hilden, Germany). Specific primers, based on the sequence for the O6-*fitD* gene, were: forward (5'-CCCTCCTTG-GCCTTGATCTG-3') and reverse (5'-CAACGCCGATATCCAACAGC-3'). Specific primers for the 16S rRNA gene of *P. chlororaphis* O6, forward (5'-GAGGAACACCAGTGGCGAAG-3') and reverse (5'-TAAACCACATGCTCCACCGC-3'), were used as an internal standard. The 50 μl mixtures were incubated at 50°C for 30 min for reverse transcription, followed by PCR for 40 cycles with denaturation at 94°C for 15 s, annealing at 55°C for 30 s, and extension at 72°C for 30 s. A Stratagene Mx3000P PCR system machine (Agilent Technologies, Waldbronn, Germany) was used. Controls involved the omission of one of the primers and analysis of the RNA without reverse transcription. Data were analyzed using the software provided by the manufacturer. The image is representative of three independent studies.



Supplementary Fig. 2. Colonization of tomato roots by *Pseudomonas chlororaphis* O6 wild-type (wt) or the *hcnA* mutant (Δ*hcnA*). Tomato seeds (TENTEN, Koregon, Korea) were surface sterilized with 70% ethanol for 5 min, treated with 1% sodium hypochlorite for 1 min and rinsed three times in sterile distilled water. Seedlings were transferred into sterilized growth mix in Magenta-boxes (7.2 × 7.2 × 10 cm) and incubated under conditions of 16 h light and 8 h dark at 26°C. The roots of 14 day-old plants were treated with 10 ml of bacterial cells suspended in sterile water and adjusted to OD_{600nm} = 0.1. At defined times, plants were carefully removed from the pots, and roots were gently shaken to remove all but the tightly adhering soil. Roots were excised, transferred to 10 ml of sterile distilled water and serial dilutions were plated onto to KB medium. The plates were incubated at 28°C for 24 h for colony counts. The number of bacteria colonizing the root was calculated as colony forming units (cfu)/g of root. The means of two separate experiments each with three tomato seedling/replicate and standard deviations (vertical bars) are provided.



Supplementary Fig. 3. Hydrogen cyanide production from the tomato rhizosphere when roots were grown with inocula of *Pseudomonas chlororaphis* O6 wild-type (wt), the *hcnA* mutant (Δ*hcnA*), the complemented *hcnA* mutant (*ComΔhncA*), or the *gacS* mutant (Δ*gacS*). Tomato seeds (TENTEN, Koregon, Korea) were surface sterilized with 70% ethanol for 5 min, treated with 1% sodium hypochlorite for 1 min and rinsed three times in sterile distilled water. Seeds were transferred to sterilized soil in Magenta-boxes (7.2 × 7.2 × 10 cm) and incubated under conditions of 16 h light and 8 h dark at 26°C. Two weeks after seeding the roots were drenched with 10 ml of bacterial cells suspended in sterile water (1×10^8 cfu/ml) or with KB broth medium (negative control). Three-days after root drenching, hydrogen cyanide production was determined qualitatively using Cyantesmo paper (Machery-Nagel GmbH & Co., Duren, Germany). The image shown represents three independent experiments, with similar results.